

This Page Is Inserted by IFW Operations  
and is not a part of the Official Record

## **BEST AVAILABLE IMAGES**

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

**IMAGES ARE BEST AVAILABLE COPY.**

**As rescanning documents *will not* correct images,  
please do not report the images to the  
Image Problems Mailbox.**

**THIS PAGE BLANK (USPTO)**

## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification<sup>5</sup>:

C12N 15/62, A61K 48/00, C12N 15/87

A1

(11) International Publication Number:

WO 94/25608

(43) International Publication Date: 10 November 1994 (10.11.94)

(21) International Application Number: PCT/US94/04589

(22) International Filing Date: 25 April 1994 (25.04.94)

(30) Priority Data:

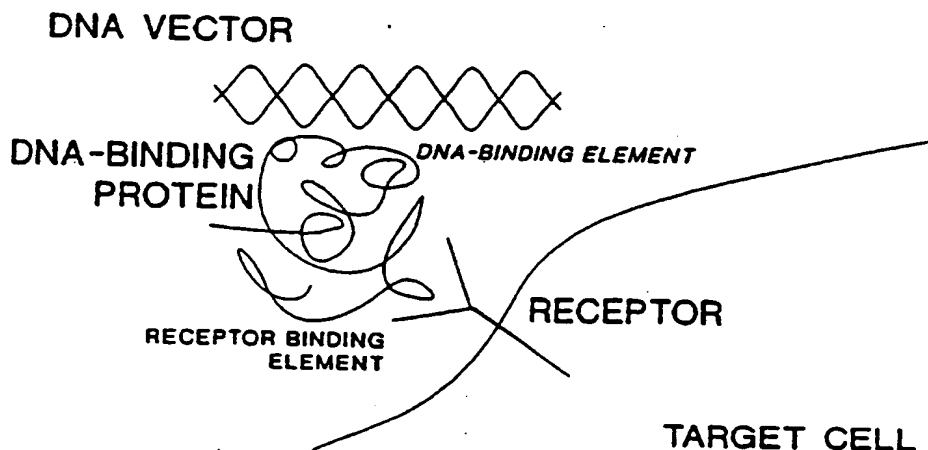
08/054,493

27 April 1993 (27.04.93)

US

(71) Applicant: BAYLOR COLLEGE OF MEDICINE [US/US];  
Texas Medical Center, One Baylor Plaza, Houston, TX  
77030-3498 (US).(72) Inventors: LEDLEY, Fred, D.; 4911 Braesvalley, Houston, TX  
77096 (US). STANKOVICS, Jozsef; Bem u.2, H-7621 Pecs  
(HU).(74) Agents: WEISS, Steven, M. et al.; Lyon & Lyon, 34th floor,  
611 West 6th Street, Los Angeles, CA 90017 (US).(81) Designated States: AT, AU, BB, BG, BR, BY, CA, CH, CN,  
CZ, DE, DK, ES, FI, GB, GE, HU, JP, KP, KR, KZ, LK,  
LU, LV, MD, MG, MN, MW, NL, NO, NZ, PL, PT, RO,  
RU, SD, SE, SK, UA, UZ, VN, European patent (AT, BE,  
CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT,  
SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML,  
MR, NE, SN, TD, TG).

## Published

*With international search report.**Before the expiration of the time limit for amending the  
claims and to be republished in the event of the receipt of  
amendments.*(54) Title: NATURAL OR RECOMBINANT DNA BINDING PROTEINS AS CARRIERS FOR GENE TRANSFER OR GENE  
THERAPYUSE OF DNA-BINDING PROTEIN FOR  
TRANSFER OF DNA INTO TARGET CELLS

## (57) Abstract

A complex for gene transfer including a DNA molecule specifically and non-specifically bound to a DNA-binding protein. Additionally, it can include a chimeric compound for gene transfer. The chimeric compound has a DNA-binding element and a ligand binding element. The chimeric recombinant DNA can also include a binding protein which has a first element for binding to a receptor, a second element for binding to DNA, a third element for destabilizing endosomes and a fourth element for directing the traffic in a protein containing complex in the nucleus of a cell. The complex will be used for the treatment of a variety of diseases.

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AU	Australia	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgyzstan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

DESCRIPTIONNatural or Recombinant DNA Binding Proteins as  
Carriers for Gene Transfer or Gene TherapyField of the Invention

The present invention relates generally to the use of DNA binding proteins to effect or enhance entry of DNA into cells for gene transfer or gene therapy. It more particularly relates to the production of chimeric, recombinant, or synthetic proteins containing DNA binding elements for the purposes of effecting or enhancing gene transfer or gene therapy. It also relates to the use of lactoferrin for enhancing entry of DNA into cells.

10 Background of Invention

The essential process of somatic gene therapy is the ability to perform gene transfer. In this process, recombinant genes are introduced into selected somatic cells. In its simplest form, gene transfer can be performed by simply injecting minute amounts of DNA into the nucleus of a cell. This process is called microinjection (1). Once recombinant genes are introduced into a cell, they can be recognized by the cells normal mechanisms for transcription, and a gene product will be expressed. The utility of microinjection for gene therapy is obviously limited by the number of cells which can be injected. Thus, more efficient methods have been developed for introducing DNA into larger numbers of cells. These methods include: transfection, where DNA is precipitated with  $\text{CaPO}_4$  and taken into cells by pinocytosis (2); electroporation, where cells are exposed to large voltage pulses to introduce holes into the membrane (3); lipofection/liposome fusion, where DNA is packaged into lipophilic vesicles which fuse with a target cell (4); and particle bombardment using DNA bound to small projectiles (5). Another important method for introducing DNA into

cells is to couple the DNA to chemically modified proteins. These modified proteins have the ability to bind DNA through a chemically attached synthetic polylysine peptide, and bind to specific receptors on target cells.

5 After these complexes are taken up by a specific receptor mediated endocytosis, the genes encoded by the DNA can be expressed by the target cell. Experiments have been performed with transferrin/polylysine/DNA complexes (6-11) as well as with asialoglycoprotein/polylysine/DNA

10 complexes (12-25). The covalently, chemically combined natural ligands are used: (1) to specifically targeted DNA to different tissues; (2) to provide more efficient uptake process. These methods are limited because they require in vitro modification of the ligands using

15 chemical or enzymatic methods in order to create a compound capable of binding DNA.

An important advance in methods of gene transfer was the demonstration that adenovirus proteins are capable of destabilizing endosomes and enhancing the uptake of DNA

20 into cells. The admixture of adenovirus to solutions containing DNA complexes, or the binding of DNA to adenoviral particles by polylysine covalently attached to the adenovirus using protein crosslinking agents substantially improves the uptake and expression of the

25 recombinant gene (6). Further studies have also suggested that proteins purified from other pathogenic bacteria, viruses or parasites may have similar properties.

In vivo studies with asialo-orosomucoid/polylysine/DNA complexes have shown that it is

30 possible to achieve short term expression (several days) of recombinant genes in organs such as the liver (12-15). For this expression to be useful, however, it will be necessary to administer the complex repetitively either to treat intermittent symptoms of disease, or to establish

35 chronic, steady-state levels of the gene product. One problem with repetitive administration using the asialo-orosomucoid/polylysine/DNA complex method, however, is

that this complex is highly antigenic and that anaphylaxis can result from repetitive administration. Significantly, antibodies are formed against both the covalently modified orosomucoid as well as polylysine. Presumably, the natural  
5 protein (orosomucoid) which is not normally antigenic, is rendered antigenic by the covalent addition of polylysine. (It is less likely that removal of sialic acid makes this protein antigenic since this is a normal intermediate in synthesis and degradation of the protein.)

10 The present invention describes an improved method for gene transfer which enables specific targeting of DNA, enhanced uptake in various cell types and endosomal destabilization without the need to produce and purify viral proteins and without the need for covalent, chemical  
15 or enzymatic modifications to couple the DNA to the ligand. The present invention thus provides a method for gene transfer which will be easier and safer than currently available methods.

#### Literature Cited in Text

- 20 (1) Capecchi MR. High efficiency transformation by direct microinjection of DNA into cultured mammalian cells. Cell 22:479-88 (1980).
- (2) Chen C. and Okayama H. High-efficiency transformation of mammalian cells by plasmid DNA. Mol.  
25 Cell Biol. 7:2745-52 (1987).
- (3) Chu G., Hayakawa H. and Berg P. Electroporation for the efficient transfection of mammalian cells with DNA. Nucleic Acids Res. 15:1311-26 (1987).
- (4) Felgner PL., Gadek TR., Holm M., et al.  
30 Lipofection: a highly efficient, lipid-mediated DNA-transfection procedure. Proc. Natl. Acad. Sci. USA. 84:7413-7 (1987).
- (5) Yang NS., Burkholder J., Roberts B, Martinell B. and McCabe D. In vivo and in vitro gene transfer to  
35 mammalian somatic cells by particle bombardment. Proc. Natl. Acad. Sci. 87:9568-72 (1990).

- (6) Curiel DT., Agarwal S., Romer MU., Wagner E., Cotten M., Birnstiel ML. and Boucher RC. Gene transfer to respiratory epithelial cells via the receptor-mediated endocytosis pathway. *Am. J. Respir. Cell. Mol. Biol.* 5 6:247-52 (1992).
- (7) Wagner E., Cotten M., Mechtler K., Kirlappos H. and Birnstiel ML. DNA-binding transferrin conjugates as functional gene-delivery agents: synthesis by linkage of polylysine or ethidium homodimer to the transferrin carbohydrate moiety. *Bioconjug-Chem.* 2:226-31 (1991). 10
- (8) Wagner E., Cotten M., Foisner R. and Birnstiel ML. Transferrin-polycation-DNA complexes: the effect of polycations on the structure of the complex and DNA delivery to cells. *Proc. Natl. Acad. Sci. USA.* 88:4255-9 15 (1991).
- (9) Wagner E., Zenke M., Cotten M., Beug H. and Birnstiel ML. Transferrin-polycation conjugates as carriers for DNA uptake into cells. *Proc. Natl. Acad. Sci. USA.* 87:3410-4 (1990).
- (10) Zenke M., Steinlein P., Wagner E., Cotten M., Beug H. Birnstiel ML. Receptor-mediated endocytosis of transferrin-polycation conjugates: an efficient way to introduce DNA into hematopoietic cells. *Proc. Natl. Acad. Sci. USA.* 87:3655-9 (1990). 20
- (11) Cotten M. Langle-Rouault F., Kirlappos H., Wagner E., Mechtler K., Zenke M., Beug H. and Birnstiel ML. Transferrin- polycation-mediated introduction of DNA into human leukemic cells: stimulation by agents that affect the survival of transfected DNA or modulate transferrin receptor levels. *Proc. Natl. Acad. Sci. USA.* 25 87:4033-7 (1990). 30
- (12) Wilson JM., Grossman M., Wu CH., Chowdhury NR., Wu GY. and Chowdhury JR. Hepatocyte-directed gene transfer in vivo leads to transient improvement of 35 hypercholesterolemia in low density lipoprotein receptor-deficient rabbits. *J. Biol. Chem.* 267:963-7 (1992).



(13) Wilson JM., Grossman M., Cabrera JA., Wu CH., Wu GY. A novel mechanism for achieving transgene persistence in vivo after somatic gene transfer into hepatocytes. J. Biol. Chem. 267: 11483-9 (1992).

5 (14) Wu GY., Wilson JM., Shalaby F., Grossman M., Shafritz DA. and Wu CH. Receptor-mediated gene delivery in vivo. Partial correction of genetic analbuminemia in Nagase rats. J. Biol. Chem. 266:14338-42 (1991).

(15) Wu CH., Wilson JM. and Wu GY. Targeting genes:  
10 delivery and persistent expression of a foreign gene driven by mammalian regulatory elements in vivo. J. Biol. Chem. 264: 16985-7 (1989).

(16) Wu GY. and Wu CH. Receptor-mediated in vitro gene transformation by a soluble DNA carrier system. J.  
15 Biol. Chem. 262:4429-32 (1987).

(17) Wu GY. and Wu CH. Targeted delivery and expression of foreign genes in hepatocytes. Targeted Diagn. Ther. 4:127-49 (1991).

(18) Wu GY. and Wu CH. Delivery systems for gene  
20 therapy. Biotherapy 3:87-95 (1991).

(19) Wu GY. and Wu CH. Receptor-mediated gene delivery and expression in vivo. J. Biol. Chem. 263:14621-4 (1988).

(20) Wu GY. and Wu CH. Evidence for targeted gene  
25 delivery to Hep G2 hepatoma cells in vitro. Biochemistry 27:887-92 (1988).

#### Summary of Invention

An object of the present invention is the use of natural DNA-binding proteins to enhance the entry of DNA  
30 into cells.

An additional object of the present invention is the use of natural DNA-binding proteins bound to DNA for the purposes of gene transfer and somatic gene therapy.

A further object of the present invention is the use  
35 of lactoferrin as a DNA-binding protein for the purposes

of effecting or enhancing the entry of DNA into cells for the purposes of gene transfer or gene therapy.

Another object of the present invention is the use of recombinant DNA-binding proteins to enhance the entry of  
5 DNA into cells.

A further object of the present invention is the use of recombinant DNA-binding proteins bound to DNA for the purposes of gene transfer and somatic gene therapy.

An additional object of the present invention is the  
10 design of chimeric proteins containing elements that bind to DNA and elements which destabilize the endosome.

An additional object of the present invention is the design of chimeric proteins containing elements which effect the trafficking of the DNA or DNA-protein complex  
15 out of the endosome.

An additional object of the present invention is the design of chimeric proteins containing elements that bind to DNA and elements which direct the trafficking of the DNA or DNA-protein complex to the nucleus.

20 Thus in accomplishing the foregoing objects, there is provided in accordance with one aspect of the present invention a complex for gene transfer comprising a DNA molecule bound to a DNA-binding protein.

In one embodiment of the present invention the DNA-  
25 binding protein binds nonspecifically to single strand, double strand, circular, or supercoiled molecules comprised of deoxyribonucleic acid.

In an embodiment of the present invention the DNA-binding protein may bind specifically to particular  
30 nucleotide sequences incorporated within the DNA.

In an additional embodiment of the present invention, the DNA-binding protein is a chimeric protein containing an element that binds to DNA and a ligand element which binds to specific receptors on target cells.

- 5 In one embodiment of this invention, lactoferrin constitutes both the DNA-binding elements and the ligand elements.

In specific embodiments of this invention, the DNA-binding element is derived from the set of genes encoding  
10 proteins including lactoferrin, histones, steroid hormone receptors, trans acting regulatory elements, basic nuclear proteins, or chromatin elements; and the ligand element is derived from the set of genes encoding proteins including  
15 protein hormones, glycoprotein hormones, serum proteins, vitamin binding proteins, transcobalamin I, transcobalamin II, R binder, intrinsic factor, cell surface proteins, cytokines, interleukins, interferons, neurotropic peptides, viral proteins, bacterial proteins or cell adhesion molecules.

- 20 In another embodiment of this invention, an additional element which effects trafficking out of the endosome is used. This element is derived from the set of genes encoding proteins including elements of adenovirus, parainfluenza virus, proteins from viruses, proteins from  
25 parasites, and proteins from bacteria.

In a specific embodiment of this invention, an element which fuses to cell membranes including the hemagglutinin of sendai virus and influenza virus are used.

- 30 Another embodiment of the present invention includes an element which directs trafficking of proteins or protein complexes to the nucleus. This element is derived from the set of gene encoding proteins including nuclear proteins from mammals, nuclear hormone receptors, viral  
35 proteins, non-viral proteins.

In an additional embodiment of this invention the DNA-binding protein is a riboprotein. The riboprotein can

bind to the DNA by hybridization or by triplex formation between the ribonucleotide sequence on the riboprotein and the DNA.

One specific embodiment of the present invention comprises a chimeric recombinant DNA-binding protein. The DNA-binding protein is comprised of elements derived from the set including: a first element for binding to a receptor on the target cell, a second element for binding to DNA, a third element capable of effecting trafficking out of the endosome or fusing with membranes, and a fourth element capable of directing trafficking of proteins or protein containing complexes to the nucleus of the cell. These elements are combined in tandem fashion to comprise a single protein molecule. These elements are combined in an order, spacing, and orientation appropriate for their multiple and specific functions.

Other and further objects, features and advantages will be apparent from the following descriptions of the presently preferred embodiments in the invention which are given for the purpose of disclosure and when taken in conjunction with the accompanying drawings.

#### Brief Description of the Drawings

##### Figure Legends

Figure 1 compares the previous schemes for gene therapy with the scheme of the present invention. The previous schemes for gene therapy required covalent modification of a ligand by coupling the ligand to polylysine. A complex could then be formed with the negative charge of DNA binding to the positive charge of polylysine. The present scheme describes the use of proteins with DNA-binding activity, thus obviating any requirement for covalent modification of a ligand for use in gene transfer.

Figure 2 shows a schematic of the use of a DNA-binding protein for transfer of DNA into target cells.

Figure 3 shows the construction of recombinant DNA-binding protein for gene transfer.

Figure 4 shows an alternative construction of recombinant-DNA-binding protein for gene transfer.

5 Figure 5 shows binding of lactoferrin to plasmid DNA.

Figure 6 shows binding of lactoferrin to linear DNA.

Figure 7 shows protection of DNA from acid hydrolysis by lactoferrin.

10 Figure 8 shows lactoferrin mediated gene transfer into hepG2 (hepatoma) cells.

Figure 9 shows Asialo-lactoferrin mediated gene transfer into hepG2 (hepatoma) cells.

Figure 10 shows lactoferrin mediated gene transfer into sol8 cells (myoblasts).

15 The drawings are not necessarily to scale. Certain features of the invention may be exaggerated in scale or shown in schematic form in the interest of clarity and conciseness.

#### Detailed Description of the Invention

20 It will be readily apparent to one skilled in the art that various substitutions and modification may be made to the invention disclosed herein without departing from the scope and spirit of the invention.

#### Definitions

25 As used herein, "Natural DNA-binding protein" means a protein whose natural sequence is capable of binding nonspecifically to DNA or specifically to certain nucleic acid sequences found within natural or recombinant DNA molecules. Natural DNA-binding proteins may be purified  
30 from natural sources. Alternatively, natural DNA-binding proteins may be produced by recombining the gene for the natural DNA-binding protein in an expression vector capable of directing expression of the DNA-binding protein in an appropriate host cell. The protein can then be  
35 produced in a variety of cells including mammalian cells,

non-mammalian cells, insect cells, bacterial cells, yeast cells, fungal cells, or transgenic animals. It will be apparent to one skilled in the art that proteins produced in these organisms may have specific post-translational  
5 modifications but will retain their natural protein sequence.

As used herein "Recombinant DNA-binding protein" means a protein whose natural sequence has been altered using recombinant DNA methods. Alterations may include  
10 eliminating segments of the natural sequence, attenuating the length of the natural sequence at the carboxyl or amino terminal ends, changing specific amino acids present in the natural sequence by altering codons encoding the protein, adding additional amino acids to the natural  
15 sequence, or creating chimeric proteins comprised of segments from several different natural or recombinant proteins. The recombinant DNA-binding proteins are created using recombinant DNA, cloning, or amplification methods. In these procedures, genetic sequences are constructed  
20 which encode the desired DNA-binding protein with a genetic element which will direct expression of the recombinant protein in a desired host. The recombinant proteins can be produced in a variety of hosts including mammalian cells, non-mammalian cells, insect cells,  
25 bacterial cells, yeast cells, fungal cells, or transgenic animals.

As used herein a "DNA-binding element" can be DNA-binding protein, a recombinant DNA-binding protein, chimeric proteins or other natural molecules whose  
30 sequence is capable of binding nonspecific DNA specifically to certain nucleic acid sequences found within natural or recombinant DNA molecules.

As used herein "Chimeric proteins" means proteins containing sequences from two or more different natural or  
35 recombinant proteins. Chimeric proteins are produced using recombinant DNA methods.

A recombinant gene can be assembled which encodes a chimeric protein that has both a DNA-binding domain and a receptor binding domain. A complex containing this chimeric protein will bind both the DNA vector and the  
5 receptor on the target cell leading to uptake and expression of the recombinant Gene (see Figure 3).

Further, a recombinant gene can be assembled which encodes a chimeric protein that has a DNA-binding domain, a receptor binding domain, and a domain capable of  
10 altering intracellular trafficking of the complex (i.e. exit from the endosome or entry into the nucleus. A complex containing this chimeric protein will bind both to the DNA vector and the receptor on the target cell leading to uptake of DNA into the cell. Once within the cell the  
15 trafficking of the complex is enhanced by the presence of a domain which alters the rate of exit from the endosome or entry into the nucleus (see Figure 4).

As used herein "Recombinant DNA methods" includes the process of separating fragments of DNA, joining fragments  
20 of DNA, synthesizing fragments of DNA, or copying fragments of DNA or RNA, amplifying fragments of DNA, or any other manipulations of DNA or RNA undertaken for the purposes of producing a protein capable of binding DNA and effecting or enhancing gene transfer or gene therapy. This  
25 may involve the use of chemical modifications or enzymes including restriction endonuclease, exonuclease, endonuclease, ligase, DNA polymerase, methylase, demethylase or RNA polymerase. It should be emphasized that these modifications take place at the level of DNA  
30 rather than protein. The present invention is thus fundamentally different from the methods of the prior art.

As used herein Vector means a DNA molecule comprised of single strand, double strand, circular, or supercoiled DNA. The vector is comprised of the following elements  
35 linked sequentially at appropriate distances for allowing functional gene expression: a promotor, a 5' mRNA leader sequence, a transcription initiation site, a nucleic acid

cassette, a 3' untranslated region, and a polyadenylation site. One or more of these elements can be eliminated for specific applications. The nucleic acid cassette can include a restriction site for insertion of the nucleic acid sequence to be expressed. In a functional vector the nucleic acid cassette contains the nucleic acid sequence to be expressed including translation initiation and termination sites,

As used herein "Ligand" means a natural or recombinant protein capable of binding to a specific or nonspecific receptor associated with a target cell.

As used herein "Carrier" means a molecule bound to DNA which effects or enhances the process of gene transfer.

As used herein "DNA-protein complexes" means the complex formed between a DNA molecule or vector and a natural or recombinant DNA-binding protein.

As used herein "Gene transfer" means the process of introducing a foreign DNA molecule into a cell. Gene transfer is commonly performed for the purposes of expressing a particular product encoded by the gene. The product may include a protein, polypeptide, anti-sense DNA or RNA, or enzymatically active RNA. Gene transfer can be performed in cultured cells or by direct administration into animals. Generally gene transfer involves the process of DNA contact with a target cell by non-specific or receptor mediated interactions, uptake of DNA into the cell through the membrane or by endocytosis, and release of DNA into the cytoplasm from the plasma membrane or endosome. Expression may require, in addition, movement of the DNA into the nucleus of the cell and binding to appropriate nuclear factors for transcription.

As used herein "Gene therapy" is a form of gene transfer and is included within the definition of gene transfer as used herein and specifically refers to gene transfer to express a therapeutic product from a cell in vivo or in vitro. Gene transfer can be performed ex vivo



on cells which are then transplanted into a patient, or can be performed by direct administration of the DNA or DNA-protein complex into the patient.

As used herein, "cytokine" refers to a protein or  
5 glycoprotein which acts on cells of the immune system to regulate the immune response.

As used herein, "neuropeptide" refers to a protein or glycoprotein which acts on cells of the central or peripheral nervous system to regulate neurological  
10 development or activity.

As used herein, "receptor" refers to a molecule within a cell that interacts with an exogenously applied substance and mediates an effect of that substance on the cell.

15 The present invention describes an improved method for gene transfer. A specific embodiment is a complex for gene transfer comprising a DNA molecule specifically or non-specifically bound to a DNA-binding protein. The DNA-binding protein may be selected from naturally occurring  
20 proteins or the DNA-binding protein can be produced by recombinant methods which create a new, chimeric protein or protein with an altered amino acid sequence.

One skilled in the art readily recognizes that by identifying or constructing a protein which is  
25 autonomously capable of binding DNA, a DNA-protein complex can be assembled without further chemical or enzymatic modifications. If this DNA-binding protein has, in addition, elements which may effect or enhance the process of gene transfer, then this DNA-binding protein may be  
30 used to effect or enhance the process of gene transfer or gene therapy. This is without the need for chemical covalent or enzymatic modifications of the protein and provide significant advantage in manufacturing and quality control.

35 Another embodiment of the present invention is a chimeric recombinant DNA-binding protein comprising a first element for binding to a receptor on a target cell

and a second element required for binding to DNA. Further enhancements can include a third element capable of directing the trafficking of DNA out of the endosome; and/or a fourth element capable of directing the traffic  
5 in a protein or protein complex to the nucleus of a cell. The various combination of the above four elements are combined to comprise a single protein molecule and the order, space and orientation of the elements is appropriate for the multiple and specific functions of the  
10 molecule. These chimeric proteins are designed to contain elements capable of effecting several different processes required for gene transfer. The specific element can be selected from: (1) ligands for specific or nonspecific receptors on the surface of a cell, thus bringing DNA into  
15 contact with the cell; (2) fusion of the DNA-protein complex with the cell membrane; (3) endocytosis of the DNA-protein complex, destabilization of the endosome leading to release of the DNA-protein complex into the cell; and (4) trafficking of the DNA-protein complex to  
20 the nucleus.

The chimeric protein is constructed by recombining gene sequences which encode the desired elements into a single DNA molecule and expressing the product of the recombinant gene in an appropriate host.

25 In the chimeric protein these elements are positioned in a single linear sequence with the elements positioned in an order, spacing, and orientation that is appropriate for the desired activity.

Any one of the DNA-binding elements can bind  
30 nonspecifically to different forms of DNA including single strand, double strand, linear, circular, or supercoiled or may bind specifically to particular sequences within the sequence. In this embodiment, the bond between the DNA-binding protein and DNA will be non-covalent.

35 The elements which are ligands for specific or nonspecific receptors are selected based on their ability to bind to the desired target cell with appropriate

affinity. This binding will bring the DNA-protein complex into contact with the cell which is the first step in gene transfer. These elements can be selected from the set including proteins, hormones, glycoprotein hormones, serum  
5 proteins, secreted proteins, cell surface proteins, cytokines, interleukins, interferons, neurotropic peptides, viral proteins, or bacterial proteins, cell adhesion molecules, immunoglobulins, T-cell receptors, cell surface markers from mature or immature bone marrow  
10 elements, or cell surface markers from lymphocytes.

In the present invention a DNA-binding protein containing both a DNA-binding element and a receptor binding element (ligand) is used for gene transfer into target cells. A complex containing the DNA-binding protein  
15 and the DNA vector will bind to receptors on the target cell leading to uptake and expression of the recombinant gene. In the preferred embodiment of this invention the DNA-binding protein is lactoferrin.

One skilled in the art will recognize that it is  
20 possible to select the ligand element to direct gene transfer or gene therapy to different cell types. For example, IL2 for T-cells; somatostatin for pancreatic B-cells; endorphin for regions of the brain, and MSH for melanocytes.

25 Elements which cause fusion of the DNA-protein complex with the cell membrane will cause the DNA bound to this complex to be introduced into the cytoplasm of the cell. These elements can be selected from the set including the hemagglutinin of sendai virus, parainfluenza  
30 virus, and HIV.

The elements causing endocytosis of the DNA-protein complex are related to the elements which are ligands for various receptors in that the interaction of some ligands with their receptor is the stimulus for endocytosis. Thus  
35 the selection of the ligand will determine whether the DNA-protein complex is taken up by endocytosis into the targeted cell.

One skilled in the art will recognize that by incorporating elements capable of fusing with the cell membrane or elements capable of causing endocytosis, the method by which DNA is taken up by the cell can be selected. The efficiency of this gene transfer process can be enhanced by optimizing the structure of the chimeric DNA-binding protein.

The elements that effect trafficking of materials out of the endosome or cause destabilization of the endosome leading to release of the DNA-protein complex are derived from viral, bacterial or parasitic proteins for which endosomal destabilization is known to be an essential part of the viral life cycle. This well-known property of adenovirus or other viruses prevents the viral particle from being destroyed during the process of infection. It has been shown that the presence of adenoviral particles during the process of receptor mediated endocytosis greatly enhances the efficiency of uptake of intact DNA. The construction of chimeric proteins containing specific genetic elements known to cause endosomal destabilization enhances the process of gene transfer.

The elements directing trafficking of the DNA-protein complex to the nucleus of the cell are derived from proteins which are normally synthesized on cytoplasmic ribosomes and then directed to the nucleus by specific sequences within the protein. These elements have been described on many proteins and have been shown to direct the trafficking of recombinant proteins to the nucleus. These elements can be derived from the set including nuclear proteins from mammals, nuclear hormone receptors, viral proteins and non-viral proteins.

One skilled in the art will recognize that transcription or replication of DNA occurs in the nucleus. Thus the presence of a nuclear localizing sequence enhances the efficiency of gene expression after the process of gene transfer.

The natural or recombinant DNA-binding protein may undergo post-translational modifications after synthesis on cytoplasmic ribosomes. Thus the protein may be a riboprotein, glycoprotein, phosphoprotein, or lipoprotein.

5 One skilled in the art will recognize that recombinant proteins expressed in different cell types can undergo different post-translational modifications. Thus expression of a natural DNA-binding protein in different cells can produce a product which has the natural protein

10 sequence but different post-translational modifications. For example, expression of a glycoprotein in a cell which is incapable of adding sialic acid to the carbohydrate will produce an asialoglycoprotein.

Proteins with different post-translational

15 modifications can have properties which are beneficial for gene transfer or gene therapy. For example, asialoglycoproteins will bind to a specific receptor on hepatic cells and DNA-asialoglycoprotein complexes have been shown to be taken up after this interaction. Thus

20 DNA-binding glycoproteins produced in a sialotransferase deficient cell are useful to target gene delivery to the liver without further chemical or enzymatic modifications.

Complexes between natural or recombinant DNA-binding proteins can be administered by any conventional

25 parenteral route including intravenous administration, injection directly into organs or somatic structures, intramuscular injection, intradermal injection, subcutaneous injection, topical administration and oral administration.

30 The use of lactoferrin as a DNA-binding protein provides a method of oral gene therapy. Lactoferrin is relatively resistant to acid hydrolysis in the stomach as well as the action of digestive enzymes and bases in the intestines. Lactoferrin is also protected from degradation

35 in the stomach and small intestines by being incorporated into lipid micelle within milk (the cream layer). The

admixture of elements of milk to DNA-lactoferrin complexes can protect the DNA-protein complex from degradation. Complexes entering the small bowel bind to the lactoferrin receptor on these cells leading to efficient gene transfer.

The following examples are offered by way of illustration and are not intended to limit the invention in any manner.

#### Example 1

##### 10 Formation of Lactoferrin-DNA Complexes for Gene Transfer

Lactoferrin will bind linear, circular, or supercoiled DNA in solution and form a complex which can be assayed by a gel retardation assay in which the migration of DNA decreases as the size of the protein/DNA complex increases (figure 5-6). Several different forms of this complex are apparent on gel retardation assays depending upon the stoichiometry of association between the DNA and lactoferrin molecules. At low concentrations of lactoferrin there is a minimal retardation of DNA migration in the gel suggesting that the complex contains few copies of lactoferrin. At high concentrations of lactoferrin the complex does not enter the gel suggesting that a larger complex, containing many copies of lactoferrin, has been formed. The complex is stable over a wide range of salt concentrations (0-1 mM) salt, and pH (pH 2-12) (figure 7). Of particular significance is the fact that at low pH, DNA is apparently protected from hydrolysis while it is part of the DNA complex (figure 7).

DNA is incubated with lactoferrin at different concentrations and then run an agarose gel. The formation of a lactoferrin-DNA complex is monitored by the retardation of DNA migration into the gel. With higher concentrations of lactoferrin there is progressive retardation of DNA migration indicating that the size and composition of the particle can be specifically altered (see Figure 5).

DNA is incubated with lactoferrin after digestion with a restriction enzyme to produce linear fragments and end labeled with [<sup>32</sup>P]-dCTP to allow visualization after gel electrophoresis. The formation of lactoferrin-DNA complex is monitored by the retardation of DNA migration into the gel. With higher concentrations of lactoferrin there is progressive retardation of DNA migration indicating that the size and composition of the particles can be specifically altered (see Figure 6).

Complexes of DNA and lactoferrin were incubated in buffers at different pH between 2.5 and 7.0 (top). Control experiments were performed under identical conditions without lactoferrin (bottom). Hydrolysis of the DNA is evident by the appearance of lower bands representing linear (broken) DNA at pH 2.5 and pH 3.0 in the lower panel. In contrast the lactoferrin DNA complex remains intact (see Figure 7).

#### Example 2

##### Lactoferrin-DNA Complexes for Oral Gene Therapy

Lactoferrin is normally absorbed from the gut via the epithelial cells. It is normally protected from degradation in the stomach and small intestines by its resistance to acid and protease as well as by forming a complex with lipid micelles in the cream layer of milk. There are several pathways by which lactoferrin may be taken up after oral administration. It may be taken up by receptor mediated endocytosis into the endosome, or it may pass through the intestinal barrier by a transcytotic pathway and enter the general circulation.

For oral gene therapy the lactoferrin DNA-complex is administered orally by itself or in a formulation containing lipids derived from, or similar to, those in the cream layer of milk. DNA bound to the lactoferrin complex is protected from degradation in the stomach and proximal small bowel and pass through the small bowel without dissociation. The complexes reaching the small

bowel bind to the lactoferrin receptor in the cells lining the bowel and enter the cells by receptor mediated endocytosis.

### Example 3

#### 5 Parenteral Therapy With Lactoferrin DNA Complexes

Many cell types have receptors for lactoferrin. Parenteral (intravenous) administration of the lactoferrin-DNA complex leads to uptake of this complex in a variety of cell types that have the lactoferrin receptor  
10 and are in contact with the vascular space. This provides a relatively non-specific means for introducing DNA into a variety of cells *in vivo*. Tissue specific gene expression is attained by employing vectors with tissue-specific promoters such that the therapeutic gene is only  
15 expressed in cells permissive to that promoter and not other cells which have the lactoferrin receptor.

### Example 4

#### Inhalation Therapy With Lactoferrin-DNA Complexes

Lactoferrin-DNA complexes can be administered by  
20 inhalation with use of a nebulizer or direct instillation through an endotracheal tube. This complex will bind to receptors on epithelial cells or alveolar cells lining the airways and enhance the efficiency of gene transfer. Large complexes may be endocytosed specifically by alveolar  
25 macrophages enabling efficient gene transfer to these cells.

### Example 5

#### Hepatic Gene Transfer With Asialolactoferrin

Liver cells contain a highly specific, high affinity  
30 receptor for asialoglycoproteins. By exposing a liver cell to the complex comprised of asialolactoferrin and DNA, this complex will bind to the asialoglycoprotein receptor and be taken into the cell by the process of receptor mediated endocytosis.



Lactoferrin is a glycoprotein which normally contains carbohydrate elements containing terminal sialic acid. Asialolactoferrin may be constructed using several methods. One involves treating native lactoferrin with the  
5 enzyme sialidase to remove terminal sialic acids. The other is to introduce a recombinant cDNA clone for lactoferrin into a CHO cell line deficient in the enzyme required for adding sialic acid to glycoproteins (sialotransferase), or other eukaryotic or prokaryotic  
10 hosts similarly lacking this enzymatic function. Such hosts will produce asialo-lactoferrin which requires no subsequent modification. This molecule has been shown to exhibit DNA-binding properties indistinguishable from native lactoferrin.

15 Asialo-lactoferrin has been shown to effect gene transfer in cultured hepatoma cells (hepG2) with efficiencies equal to, or greater than, native lactoferrin-DNA complexes or other methods requiring covalent coupling and modification of proteins such as  
20 orosomucoid. These coupled and covalently modified compounds have been shown to be effective vehicles for gene transfer into the liver in cell culture and experimental animals. The *in vitro* experiments indicate that asialolactoferrin-DNA complexes provide a similar  
25 method for gene delivery without the need for enzymatic or chemical modifications and less potential for antigenicity.

#### Example 6

##### Hepatic Gene Transfer Based on Particle Size

30 The size of the complex formed between lactoferrin and DNA can be varied by the ratio of lactoferrin: DNA (figure 6-7), by the addition of iron, or other means. Complexes which are injected intravenously that are too large to exit from the endothelium, but are small enough  
35 to exit the sinusoid of the liver and be preferentially

endocytosed by hepatic cells would be suitable for hepatic gene transfer.

#### Example 7

##### Enhanced Hepatic Gene Transfer Using Recombinant Asialolactoferrin

5 The specificity of hepatic gene transfer using asialolactoferrin via the asialoglycoprotein receptor may be compromised by the ability of the asialolactoferrin to bind to lactoferrin receptors on other tissues. Site  
10 specific mutagenesis is employed to disrupt the receptor binding determinants of lactoferrin while retaining the sites at which the carbohydrate is attached. The asialo-form of the mutagenized, recombinant gene product provides enhanced hepatic gene transfer since it would be less  
15 likely to bind to other sites.

#### Example 8

##### Use of Recombinant, Chimeric DNA-Binding Proteins for Gene Therapy

20 A DNA-protein complex with altered tropism is constructed by recombining genetic elements encoding a DNA-binding protein with genetic elements encoding the binding determinants of a natural ligand. These elements are combined in an order, spacing, and orientation so that the ligand elements which bind to the receptor are exposed  
25 in the DNA-protein complex.

For example, a gene fragment encoding the 5' end of the cDNA encoding lactoferrin encodes the amino terminal portion of lactoferrin where the DNA-binding determinants reside. A recombinant gene can be constructed with this  
30 genetic element and the sequences encoding IL-2 attached at the 3' end of the sequence and encoding the carboxyl terminal portion of the protein. The gene product encoded by this chimeric clone can be produced in prokaryotic or eukaryotic expression systems. The resulting gene product  
35 will bind DNA through the DNA-binding determinants from

lactoferrin and will be capable of binding to the IL-2 receptor on T-cells and many T-cell malignancies. This complex may thus be used to introduce genes into T-cells to alter the function of the immune system, eliminate  
5 cells with the HIV provirus, or kill cells arising from T-cell malignancies.

#### Example 9

##### Chimeric Proteins for Enhanced Gene Transfer

A DNA-protein complex capable of enhanced gene  
10 transfer may be constructed by recombining genetic elements encoding a DNA-binding protein with genetic elements encoding a hemagglutinin, for example, the fusogenic peptide from para-influenza virus. These DNA elements are combined in an order, spacing, and  
15 orientation so that the chimeric protein retains the DNA-binding and receptor binding determinants. The chimeric clone is introduced into eukaryotic or prokaryotic cells to produce the recombinant gene product.

The fusogenic peptide from para-influenza virus  
20 effects fusion with periplasmic membranes or endosomes. Thus the complex containing the chimeric protein with the fusogenic peptide fuses with membranes after binding to target cells, and thus bypasses endosomal uptake, or may have enhanced uptake from the endosome after endocytosis.  
25 This increases the efficiency of gene transfer.

#### Example 10

##### Chimeric DNA-Binding Proteins With Tropism for Specific Receptors (2)

A gene fragment encoding the DNA-binding sequences  
30 from lactoferrin is recombined with a gene fragment encoding the receptor binding sequences of IL-2. This complex binds to DNA through the DNA-binding domain of lactoferrin and binds to mammary cells expressing the IL-2 receptor. The use of this chimeric protein enables gene  
35 delivery to T-cells and various leukemic cells. This is

useful in the regulation of T-cell function and in treatment of T-cell malignancies.

#### Example 11

##### Lactoferrin Mediated Transfer Into HepG2 (Hepatoma) Cells

5 HepG2 is a well differentiated cell line which is commonly used as a model for hepatocytes. HepG2 cells were exposed to a complex lactoferrin and DNA encoding the enzyme chloramphenicol acetyltransferase. The identical experiment was performed using a media which also  
10 contained non-infectious adenovirus particles. The presence of CAT activity in cells treated with LF/DNA indicates that the gene has been successfully transferred into these cells (see Figure 8). There is greater activity in the presence of adenovirus which enhances the  
15 trafficking of materials out of the endosome after receptor mediated endocytosis. Control experiments (Figure 9) demonstrate that there is no CAT activity in cells treated with DNA alone, lactoferrin alone, or adenovirus alone. CAT activity was evident in control experiments  
20 treated with DNA and adenovirus.

HepG2 cells were exposed to asialo-lactoferrin-DNA complexes in which the DNA encoded chloramphenicol acetyltransferase. The presence of CAT activity in cells treated with this complex but not DNA alone indicates that  
25 the gene has been successfully transferred into these cells.

#### Example 12

##### Lactoferrin Mediated Gene Transfer Into Sol8 Cells (Myoblasts)

30 Sol8 is a continuous myoblast line that exhibits many properties of myoblasts or myocytes including differentiation in culture. Cells were exposed to a vector encoding chloramphenicol acetyltransferase under different conditions (Figure 10). CAT activity was measured in cells  
35 transfected using lipofectin™ (GIBCO) as a positive

control, in cells exposed to media alone, in cells exposed to lactoferrin-DNA complex in media containing also adenovirus particles, in cells exposed to lactoferrin-DNA alone, in cells exposed to DNA alone, and in cells exposed  
5 to DNA in media containing also adenovirus. Adenovirus in the media was highly toxic to cells. The presence of CAT activity in cells exposed to lactoferrin-DNA complexes indicates that gene transfer into these cells is as efficient as lipofectin™.

10 All patents and publications mentioned in this specification are indicative of the levels of those skilled in the art to which the invention pertains. All patents and publications are herein incorporated by reference to the same extent as if each individual  
15 publication was specifically and individually indicated to be incorporated by reference.

One skilled in the art will readily appreciate that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned, as  
20 well as those inherent therein. The present examples of DNA vectors along with the methods, procedures, treatments, molecules, and specific compounds described herein are presently representative of preferred  
25 embodiments, are exemplary, and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art which are encompassed within the spirit of the invention as defined by the scope of the claims.

Claims

1. A chimeric compound useful as an element in gene transfer comprising:
  - a DNA-binding element, said DNA-binding element  
5 selected from the group consisting of lactoferrin, histone, nuclear hormone receptor, transacting regulatory element, basic nuclear protein and chromatin element;
  - a ligand binding element, said ligand-binding element  
10 selected from the group consisting of protein hormone, glycoprotein hormone, serum protein, vitamin binding protein, transcobalmin I, transcobalmin II, R binder, intrinsic factor, cell surface protein, cytokine, neuropeptide, viral protein, bacterial protein, cell  
15 adhesion molecule, immunoglobulin, T-cell receptor, cell surface marker from mature or immature bone marrow, and cell surface marker from lymphocyte.
2. A complex for gene transfer comprising a DNA bound to the chimeric compound of claim 1.
3. A chimeric recombinant DNA-binding protein  
20 comprising:
  - a first element for binding to a receptor on a target cell; and
  - a second element required for binding to DNAwherein the first and second elements are combined to  
25 comprise a single protein molecule and wherein the order, spacing and orientation is appropriate for the multiple and specific functions.
4. The chimeric recombinant DNA-binding protein of claim 3 further comprising a third element capable of  
30 destabilizing endosomes, wherein the first through third elements are combined to comprise a single protein molecule and wherein the order, spacing and orientation is appropriate for the multiple and specific functions.

5. The chimeric recombinant DNA-binding protein of claim 3 further comprises a fourth element capable of directing the trafficking of protein or protein containing complex to the nucleus of a cell, wherein the first, second and fourth elements are combined to comprise a single protein molecule and wherein the order, spacing and orientation is appropriate for the multiple and specific functions.

6. A chimeric recombinant DNA-binding protein comprising:

a first element for binding to a receptor on a target cell;

a second element required for binding to DNA;

a third element capable of destabilizing endosomes;

and

a fourth element capable of directing the trafficking of protein or protein containing complex to the nucleus of a cell;

wherein the first through fourth elements are combined in a random fashion to comprise a single protein molecule and wherein the order, spacing and orientation is appropriate for the multiple and specific functions.

7. A complex for gene transfer comprising a DNA bound to the chimeric recombinant binding protein of claim 3, 4, 5 or 6.

8. The complex of claim 2 wherein the DNA-binding element is lactoferrin.

9. The complex of claim 7 wherein the DNA-binding element is lactoferrin.

10. A method of gene transfer comprising the step of administering a pharmacological dose of the complex of claim 2.

11. The method of claim 10 wherein the dose is administered by the method selected from the group consisting of oral, parenteral, topical and inhalation.

12. A method of gene transfer comprising the step of  
5 administering a pharmacological dose of the complex of claim 7.

13. The method of claim 12 wherein the dose is administered by the method selected from the group consisting of oral, parenteral, topical and inhalation.

10 14. A method of gene transfer comprising the step of administering a pharmacological dose of the complex of claim 8.

15 15. The method of claim 14 wherein the dose is administered by the method selected from the group consisting of oral, parenteral, topical and inhalation.

16. A method of gene transfer comprising the step of administering a pharmacological dose of the complex of claim 9.

20 17. The method of claim 16 wherein the dose is administered by the method selected from the group consisting of oral, parenteral, topical and inhalation.

25 18. A complex for gene transfer comprising a lactoferrin molecule bound to a DNA vector, wherein said vector contains a nucleic acid fragment encoding a therapeutic product.

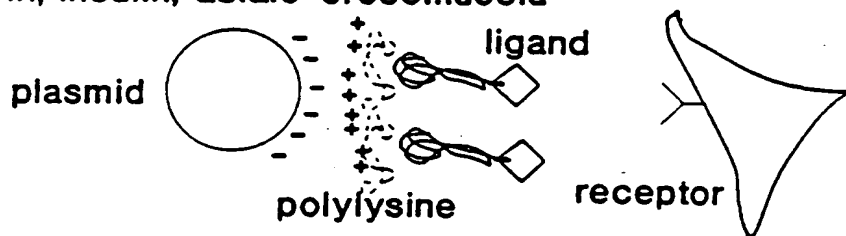
19. A method of gene therapy comprising the step of administering said lactoferrin molecule bound to a DNA vector of claim 18 to an animal.



## DNA-BINDING LIGANDS AS CARRIERS FOR DNA MEDIATED GENE TRANSFER

### Previous scheme

Transferrin, insulin, asialo-orosomucoid



### Proposed scheme

lactoferrin, recombinant chimeric proteins

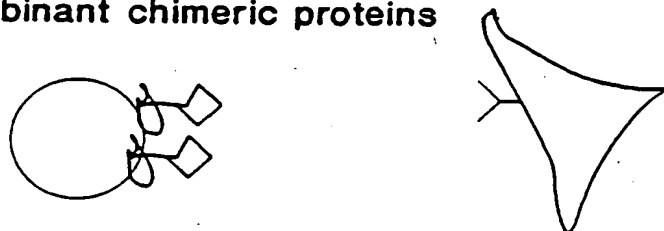


FIG. 1

# USE OF DNA-BINDING PROTEIN FOR TRANSFER OF DNA INTO TARGET CELLS

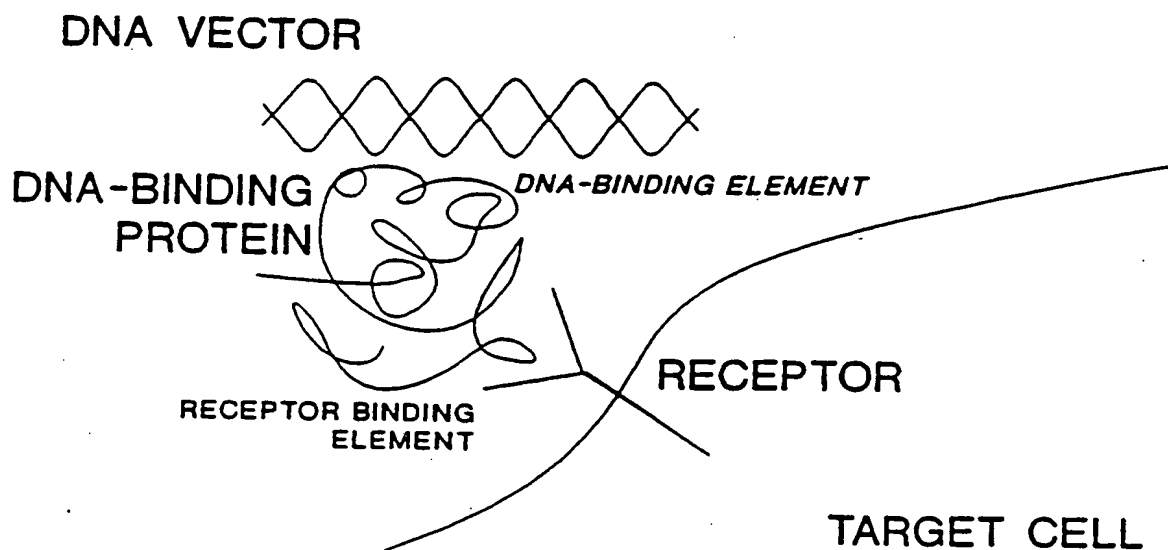


FIG. 2

# CONSTRUCTION OF RECOMBINANT DNA-BINDING PROTEIN FOR GENE TRANSFER

GENE ENCODING CHIMERIC PROTEIN

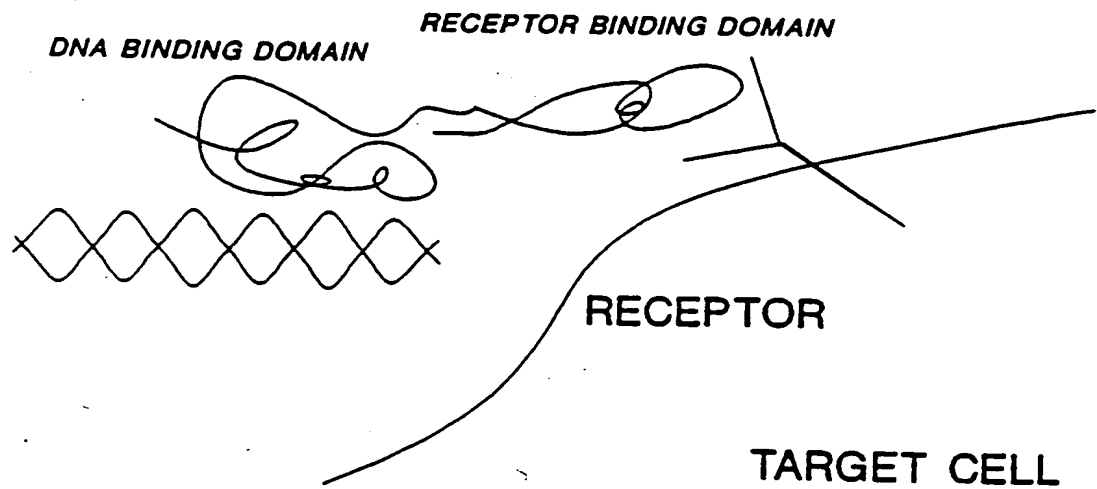


FIG. 3

## CONSTRUCTION OF RECOMBINANT DNA-BINDING PROTEIN FOR GENE TRANSFER

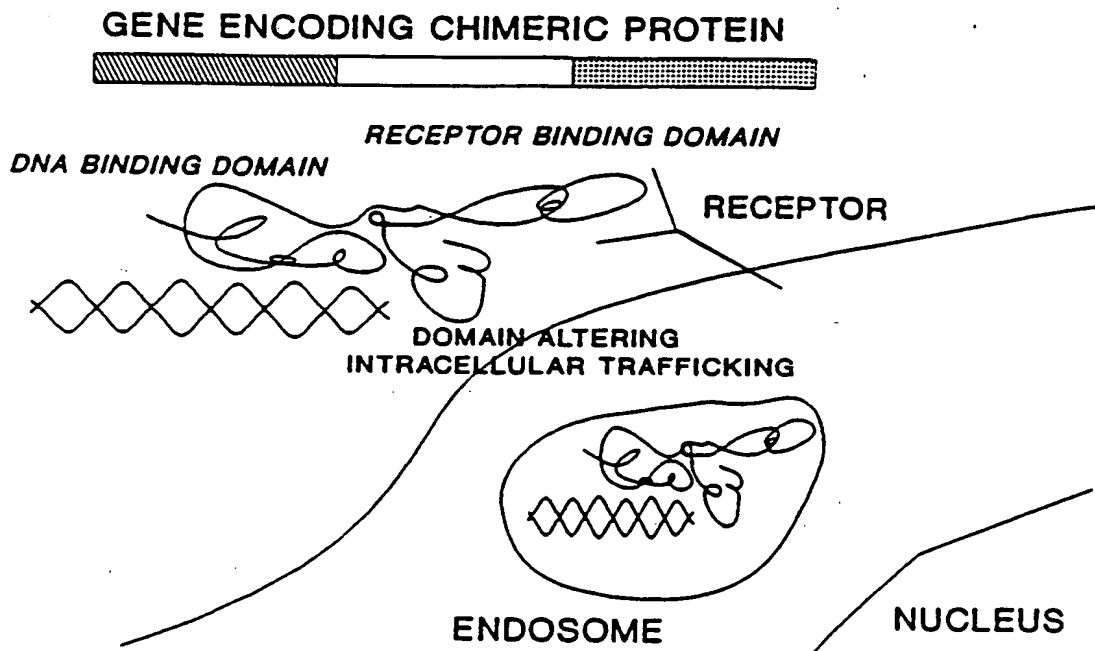


FIG. 4

## LACTOFERRIN CONCENTRATION

O 10 9 8 7 6 5 4 3 2 1 0

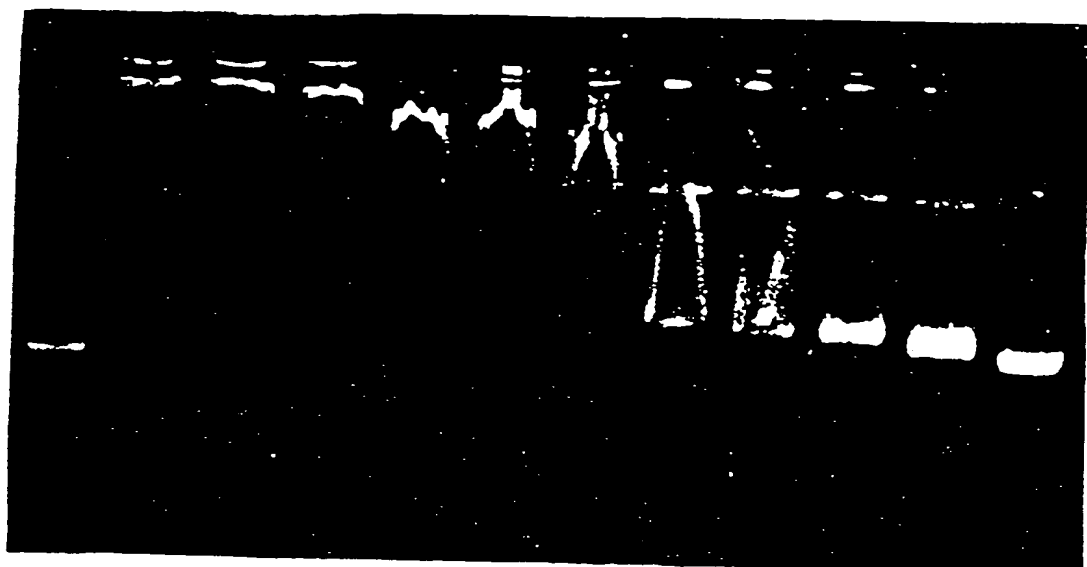


FIG. 5

## LACTOFERRIN/DNA COMPLEX

LACTOFERRIN CONCENTRATION



FIG. 6

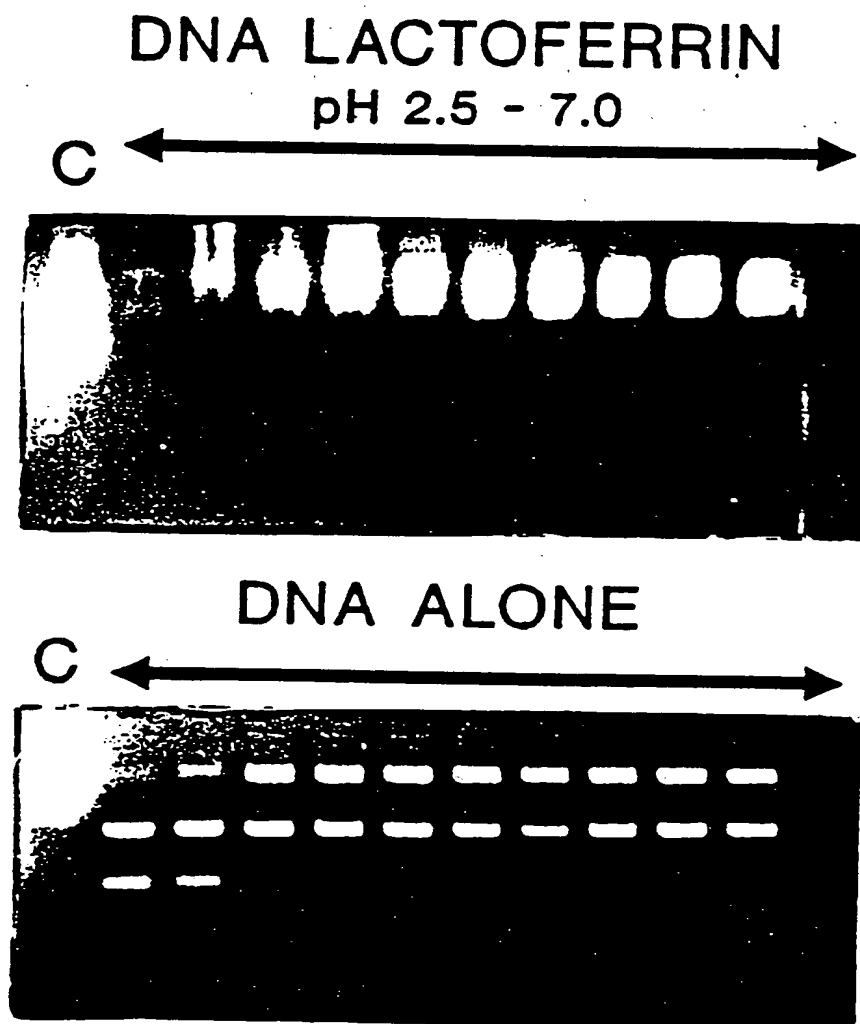


FIG. 7

# LACTOFERRIN MEDIATED GENE TRANSFER IN hepG2 CELLS

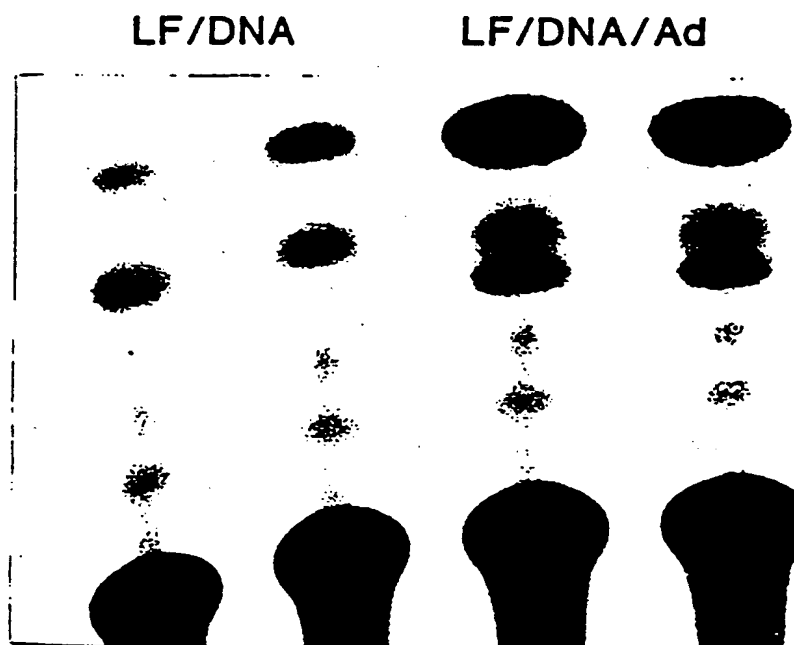


FIG. 8



ASIALO-LACTOFERRIN MEDIATED GENE TRANSFER  
INTO hepG2 CELLS

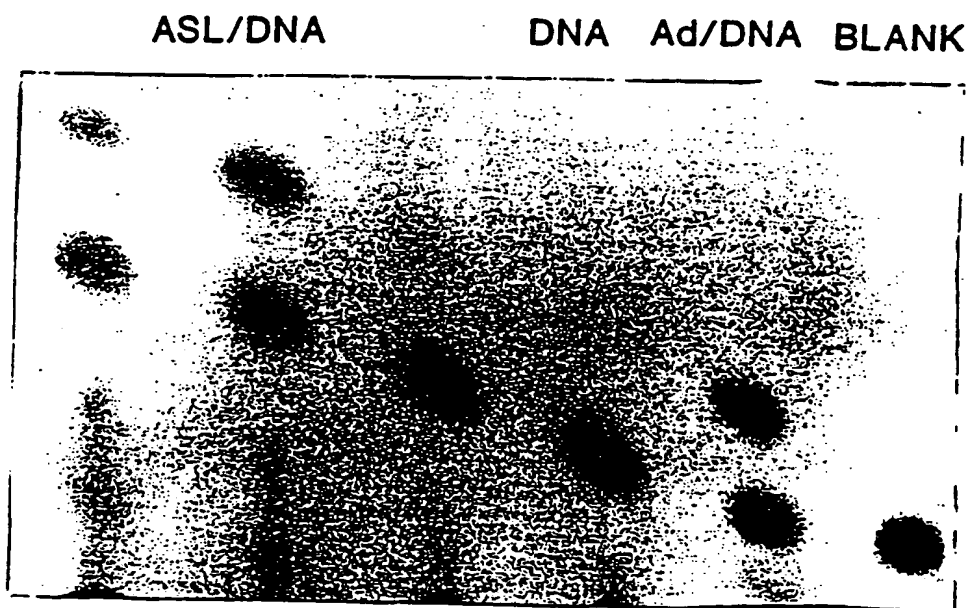


FIG. 9

## LACTOFERRIN MEDIATED GENE TRANSFER IN MYOBLASTS

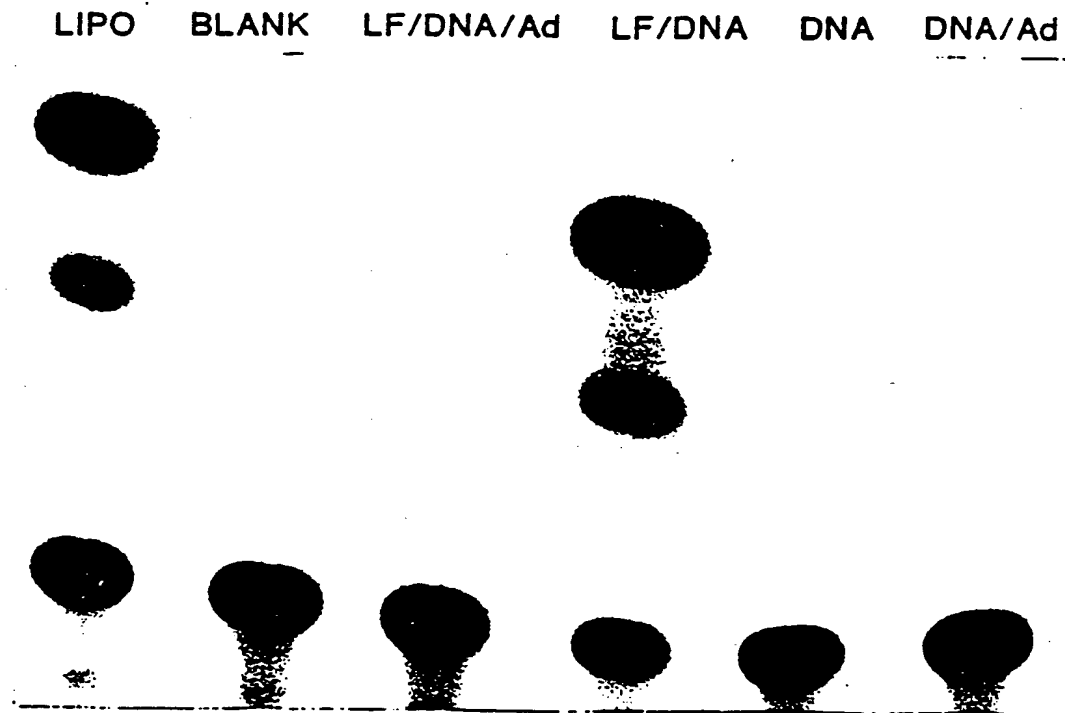


FIG. 10

A. CLASSIFICATION OF SUBJECT MATTER  
 IPC 5 C12N15/62 A61K48/00 C12N15/87

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
 IPC 5 A61K C07K C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	PATENT ABSTRACTS OF JAPAN vol. 163, no. 51 (C-0968) 29 July 1992 & JP,A,04 108 390 (SNOW BRAND MILK PROD. CO. LTD.) 1992	1-3, 8, 10, 11, 18, 19
Y	see abstract	4-7, 9
X	WO,A,92 21752 (THE UNITED STATES OF AMERICA) 10 December 1992 see claims.	1, 3
Y	J. BIOL. CHEM. vol. 268, 1993 pages 6866 - 6869 S.H. MICHAEL ET AL.; 'Binding-incompetent adenovirus facilitates molecular conjugate-mediated gene transfer by the receptor-mediated endocytosis pathway' see ABSTRACT AND INTRODUCTION.	4-7, 9
	--- -/-	

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

\* Special categories of cited documents:

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \*&\* document member of the same patent family

Date of the actual completion of the international search

19 September 1994

Date of mailing of the international search report

07. 10. 94

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
 NL - 2280 HV Rijswijk  
 Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,  
 Fax: (+ 31-70) 340-3016

Authorized officer

Yeats, S

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>J. BIOL. CHEM. vol. 264 , 1989 pages 16985 - 16987 C.H. WU ET AL.; 'Targeting genes: delivery and persistent expression of a foreign gene driven by mammalian regulatory elements in vivo' cited in the application see abstract.</p> <p style="text-align: center;">---</p>	1,3
A	<p>AM. J. RESPIR. CELL MOL. BIOL. vol. 6 , 1992 pages 247 - 252 D.T. CURIEL ET AL.; 'Gene transfer to respiratory epithelial cells via the receptor-mediated endocytosis pathway' cited in the application see abstract.</p> <p style="text-align: center;">---</p>	1,3
P,X	<p>J. CELL. BIOCHEM. SUPPL. 0 (18 PART A), ABSTRACT NO. DZ 136 1994 page 231 J. YOVANDICH ET AL.; 'Lactoferrin-mediated gene transfer: use of a nonantigenic, DNA-binding protein for receptor targeted gene therapy' &amp; Keystone Symposium on gene therapy, Copper Mountain, Colorado, USA, January 15-22, 1994.</p> <p style="text-align: center;">-----</p>	1-19



**THIS PAGE BLANK (USPTO)**